



Annona muricata EFFECT toward NITRIC-OXIDE PRODUCTION in SEVERE and ACT-TREATED-RECOVERY-PHASE of MALARIA

¹Kis Djamiatun, ²Hanif Nur Azizah

¹Parasitology Department, Faculty of Medicine Diponegoro University, Semarang, Indonesia

²PKU Hospital, Wonogiri, Indonesia.

Abstract: Chloroquine, and artemisinin reverse partly hemozoin-induced-M2 to M1-phenotype-cells in vitro. These M1-cells however, cannot significantly increase nitric oxide (NO)-production important for protecting severe-malaria-infection. An adjuvant malaria therapy might be needed to increase NO-production. The aim of this study was to determine whether water-extracted of *Annona muricata*-leaves (WAM) increased spleen-cells-NO-production of *Plasmodium berghei* ANKA (PbA)-Swiss-mice treated with Artemisinin-based-combination-therapy (ACT), an anti-malaria recommended by WHO. This study was continuing study of a post-test-only-randomized-control-group-design used 24 PbA-infected-female-Swiss-mice, divided into control-(K)-group without any intervention, and treatment-P1, P2 and P3-groups received WAM, ACT (Dihydroartemisinin Piperaquine; DHP) and WAM-ACT-combination, respectively. The spleen-cells obtained on day-7-PbA-infection were cultured. The NO-production was then measured in the cell-culture-supernatants using Griess-reagent. Parasitaemia-percentage was also evaluated. Statistical-analysis used was Kruskal-Wallis-test followed by Mann-Whitney-U-test. The NO-productions were significantly different among those four-groups ($p < 0.001$). The highest-NO-production was found in the P1-group with the severe-PbA-infection. Those of P1-group was significantly higher than K-group. Meanwhile, the lowest were found in the P2 and P3-groups with the PbA-infection-recovery-phase, and those of P2 and P3-groups were not different. The conclusion is WAM-treatment increases the splenic-NO-production during severe-PbA-infection, and no longer increases the production in the recovery-phase of those treated with ACT.

IndexTerms - *Annona muricata*, ACT, Nitric Oxide, *P. berghei* ANKA

INTRODUCTION

Plasmodium sp-infection, can develop into fatal-disease. Artemisinin-based-combination-therapy (ACT) is recommended by WHO as the main treatment for malaria cases.¹ Adjuvant therapy however is needed as a companion to anti-malarial drugs in the management of malaria cases before progressing to severe malaria, because adjuvant therapy can provide better clinical outcomes.² The immune response during malaria determines the development of severe malaria. This underlies the need to consider the provision of adjuvant therapy, which is a good immunomodulator to inhibit the development of severe malaria. The herb extensively studied as immunomodulator of experimental-cerebral-malaria (ECM), is *Annona muricata* (AM). The ethanol-extracted-(E)AM-leaves affects the spleen, an important organ for developing protective and pathologic immune responses in ECM. The EAM has anti-inflammatory, as indicated by a decrease splenic-tumour-necrosis-factor-alpha (TNF- α)-production and an increase splenic-Nitric-Oxide-(NO)-production of *Plasmodium berghei* ANKA (PbA)-infected-Swiss-mice, an ECM-susceptible-mice.³ NO involves in suppressing the parasitaemia-level of malaria-mouse-model.⁴ The treatment used is an oral-supplementation of arginine, an amino-acid involved in NO synthesis, in combination with lysine and valine, both of which are arginase-inhibitors). The treatment reduces parasitemia level which associated with increase NO-production and innate-immune-respond-dependent-MyD88-pathway. NO also protects ECM-mice from cerebrovascular-constriction.⁵ The water-extracted-AM-leaves (WAM) which is commonly consumed by the public has not been studied in detail regarding its role as an adjuvant-therapy capable of modulating splenic-NO production in severe PbA infections. In addition, ACT has immunomodulatory effect in several diseases.^{6,7} The study-aim was to determine whether WAM modulate splenic-NO-production in those ACT-treated-malaria.

MATERIALS AND METHODS

This was continuing study done before used a post-test-only-randomized with control-group-design, using 24 female-Swiss-mice which were divided into 3 treatment-groups and 1 positive-control-group.⁸ Health Research Ethical Committee Faculty of Medicine Universitas Diponegoro approved this study (Ethical Clearance No. 55/EC/H/FK-RSDK/V/2018). The sample inclusion-criteria were female-mice, weighing 20-25 grams, 6-8 weeks old, healthy, and active. The died-mice during the study, were drop-out. All samples were adapted for 6 days and randomized to K(+)-group given standard feed and distilled water, P1-group given a WAM-preventive-dose (4.68mg/ 30g mice-weight) for 7-days before and 3-days after PbA-inoculation. This then followed by a WAM-

therapeutic-dose (9.36mg/ 30g mouse-weight) until day-6-PbA-infection. P2-group was treated with ACT (0.819mg/ 30g mouse-weight) since day-4 until day-6 PbA-infection. P3-group was given both WAM and ACT. The parasitaemia-level was also observed. The mice were terminated on day-7-PbA-infection and the spleen were collected and processed for cell culture described elsewhere.⁹ The culture-supernatant of LPS-stimulated-spleen-cells was measured for NO-levels by using the Griess-reagent kit (Catalog number: 30100, Biotium, Fremont, CA, USA). Statistical analysis used was Kruskal-Wallis-test followed by Mann-Whitney-U-test with a significance degree of $p < 0.05$.

RESULTS

The Saphiro-Wilk-normality-test of NO-level showed that the data were not normally distributed. The Kruskal-Wallis showed significant different of the NO-levels among the group ($p < 0.05$), and Mann-Whitney-U-test were then carried out to assess the differences between two groups as shown in the table. The NO-production observed in this study was done on those K(+) and P1-groups with severe PbA-infection, and those P2 and P3-groups with PbA-infection-recovery-phase. The P1-group-NO-production was significantly higher than K-group. These suggested that WAM-treatment associated with an increase spleen-cell-NO-production during severe-PbA-infection. Meanwhile, those produced by P2 and P3-groups was significantly lower than K-group. The finding demonstrated that those in malaria-recovery-phase associated with a reduce-NO-production. Additionally, P2 and P3-groups showed no different of NO-production. These indicated that WAM-treatment was no longer increase NO-production in those entering the recovery-phase of those treated with ACT.

Table Mann-Whitney-U-Test of NO levels produced by spleen-cells of PbA-infected Swiss mice

Groups	NO Level Median (min – max) μmol	<i>p value</i>		
		P1	P2	P3
K (+)	40.09 (11.03 – 58.30)	0.005	0.002	0.002
P1	59.05 (44.41 – 86.07)		<0.001	<0.001
P2	11.02 (5.43 – 21.39)			0.427
P3	11.15 (8.47 – 13.96)			

DISCUSSION

WAM-treatment associated with higher spleen-NO-production of severely-PbA-infected-mice than those without treatment (Table). The culture supernatant used in this study was collected from previous study which showed WAM-treated and untreated-groups were in the severe-PbA-infection.⁸ This study therefore suggested that WAM contributed in the increase-spleen-NO-production of severe-PbA-infected-Swiss-mice. This in agreement with the findings in EAM-study using the same mouse-model.³ This also supported by the fact that phytotherapy used can lead to M1 macrophage-development.¹⁰ The lower of NO-production is observed in *P. falciparum* and PbA-infection.^{3, 11} The decrease-NO-production is due to M2-macrophage-monocyte activation in malaria-falciparum.¹¹ Monocytes become M2-type after phagocytizing the pigment *Plasmodium*-hemozoin (Hz).¹² A study using NO donors demonstrates that NO inhibits the CM-manifestation by suppressing the pathological-CD4+Th-cell and CD8+CTL of PbA-infected-mice, without influencing parasitaemia-level.¹³ Severely-PbA-infected-untreated-mice associates with a higher-brain-Granzyme-B (GzmB)-expression above normal-healthy-mice, while EAM-treatment associates with a normalize-brain-GzmB-expression in severely-PbA-infected-Swiss-mice.⁸ Whether the increase-spleen-NO-production mediated by WAM, is associated with the increase of brain-non-pathogenic CD8+T-cell-accumulation needs to be elucidated in PbA-infected-mice.

Both ACT and WAM-ACT-treated associated with lower spleen-NO-production than untreated-severe-PbA-infected-mice (Table). The spleen-NO-production difference was not observed between the two-treated-groups. The samples used in this study originated from previous study which showed those treated groups were in the recovery phase of PbA-infection with no different of parasitaemia-levels between those groups.⁸ WAM-treatment therefore, had no obvious additional effect toward spleen-NO-production in the recovery-phase of those received ACT-treatment. The findings of this study were aligned with serum-NO-levels observed in the Sago-caterpillar-*(Rhyinchophorus ferrugineus)*-flour-supplementation-study in ACT-treated-PbA-infected-Swiss-mice.¹⁴ Antimalarial *in vitro* study shows that chloroquine, and artemisinin can partially reverse Hz-phagocytizing-M2-macrophages to M1-type, indicated by no effect of those antimalaria toward arginase-activity and NO-production.¹² NO-production is induced by malaria-toxin including combination of Hz-malaria-DNA, and glycosylphosphatidylinositols (GPI).¹⁵ This therefore, the influence of both ACT and malaria toxin on NO-production needs to be further studied. The conclusion is that WAM-treatment increases NO-spleen-production in severe-PbA-infection, meanwhile the WAM limits the increase of NO-spleen-production in the recovery phase of ACT-treated-PbA-infection.

REFERENCES

1. Anonymous, Severe Malaria. Tropical Medicine & International Health, 2014; 19(s1).7-131.DOI: https://doi.org/10.1111/tmi.12313_2.
2. Varo R, Crowley VM, Siteo A, Madrid L, Serghides L, Kain KC, and Bassat Q, Adjunctive Therapy for Severe Malaria: A Review and Critical Appraisal. Malar J, 2018; 17(1).47.DOI: 10.1186/s12936-018-2195-7.
3. Maria Estela K, Fransisca Prameshinta H, Dharmana E, and Djamiatun K, Efektivitas Ekstrak Daun Sirsak (Annona Muricata) Dalam Menurunkan Kadar Tnf α Dan Meningkatkan Kadar No Uji Coba Pada Mencit Swiss Yang Diinokulasi Plasmodium Berghei Anka. Jurnal Kedokteran Brawijaya, 2016; 29.4.

4. Meireles P, Brás D, Fontinha D, Chora Â F, Serre K, Mendes AM, and Prudêncio M, Elimination of Hepatic Rodent Plasmodium Parasites by Amino Acid Supplementation. *iScience*, 2020; 23(12).101781.DOI: 10.1016/j.isci.2020.101781.
5. Ong PK, Moreira AS, Daniel-Ribeiro CT, Frangos JA, and Carvalho LJM, Reversal of Cerebrovascular Constriction in Experimental Cerebral Malaria by L-Arginine. *Sci Rep*, 2018; 8(1).15957.DOI: 10.1038/s41598-018-34249-2.
6. Bai L, Li H, Li J, Song J, Zhou Y, Liu B, Lu R, Zhang P, Chen J, Chen D, Pang Y, Liu X, Wu J, Liang C, and Zhou J, Immunosuppressive Effect of Artemisinin and Hydroxychloroquine Combination Therapy on Iga Nephropathy Via Regulating the Differentiation of Cd4+ T Cell Subsets in Rats. *International Immunopharmacology*, 2019; 70.313-323.DOI: 10.1016/j.intimp.2019.02.056.
7. Liang N, Zhong Y, Zhou J, Liu B, Lu R, Guan Y, Wang Q, Liang C, He Y, Zhou Y, Song J, and Zhou J, Immunosuppressive Effects of Hydroxychloroquine and Artemisinin Combination Therapy Via the Nuclear Factor-Kb Signaling Pathway in Lupus Nephritis Mice. *Experimental and therapeutic medicine*, 2018; 15(3).2436-2442.DOI: 10.3892/etm.2018.5708.
8. Sulayman A, Djamiatun K, and Muniroh M, Effectivity of Annona Muricata and Artemisinin Combined Therapy on Brain Cxcl10 Expression (Study in Swiss Mice During Severe Plasmodium Berghei Anka Infection). *Journal of Biomedicine and Translational Research*, 2019; 5.47-52.DOI: 10.14710/jbtr.v5i2.4802.
9. Djamiatun K, Naamat WFA, Dharmana E, Wijayahadi N, and Nugroho D, Reduce Spleen-Ifn-Gamma Correlated with Cxcl9 Levels During Cerebral Malaria Phase in Annona Muricata-Treated Swiss Mouse Study. *Advanced Science Letters*, 2017; 23(4).3380-3384.DOI: 10.1166/asl.2017.9179.
10. Saeedifar AM, Mosayebi G, Ghazavi A, Bushehri RH, and Ganji A, Macrophage Polarization by Phytotherapy in the Tumor Microenvironment. *Phytother Res*, 2021; 35(7).3632-3648.DOI: 10.1002/ptr.7058.
11. Weinberg JB, Volkheimer A, Rubach M, Florence S, Mukemba J, Kalingonji A, Langelier C, Chen Y, Bush M, Yeo T, Granger D, Anstey N, and Mwaikambo E, Monocyte Polarization in Children with Falciparum Malaria: Relationship to Nitric Oxide Insufficiency and Disease Severity. *Scientific Reports*, 2016; 6.29151.DOI: 10.1038/srep29151.
12. Bobade D, Khandare AV, Deval M, Shastry P, and Deshpande P, Hemozoin-Induced Activation of Human Monocytes toward M2-Like Phenotype Is Partially Reversed by Antimalarial Drugs-Chloroquine and Artemisinin. *Microbiologyopen*, 2018.e00651.DOI: 10.1002/mbo3.651.
13. Jeney V, Ramos S, Bergman ML, Bechmann I, Tischer J, Ferreira A, Oliveira-Marques V, Janse CJ, Rebelo S, Cardoso S, and Soares MP, Control of Disease Tolerance to Malaria by Nitric Oxide and Carbon Monoxide. *Cell Rep*, 2014; 8(1).126-136.DOI: 10.1016/j.celrep.2014.05.054.
14. Ariani, Anjani G, Adji MAS, and Djamiatun K, Tepung Ulat Sagu (Rhyinchophorus Ferrugineus) Imunomodulator Nitric Oxide (No) Sirkulasi Mencit Terapi Antimalaria Standar. *The Indonesian Journal of Nutrition*, 2018; 6(2).131 - 138.
15. Starkl Renar K, Iskra J, and Križaj I, Understanding Malarial Toxins. *Toxicon*, 2016; 119.319-329.DOI: <https://doi.org/10.1016/j.toxicon.2016.06.017>.

